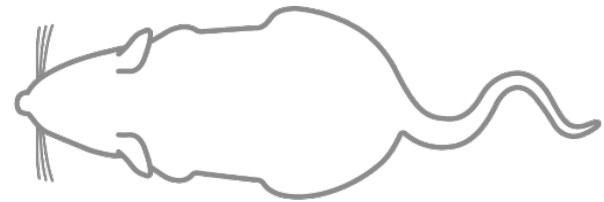


# Evidence-based predictive accuracy evaluation of the sterilizing mouse model

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September 11, 2017



# Current TB regimen development

## *Risk of late-stage attrition*

### PRECLINICAL

Varied models and approaches currently applied

### PHASE I-IIa

Safety PKPD  
Dose-Ranging PK  
14-Day EBA  
(Whole Blood Assay?)

### PHASE IIb

Dosing  
POC-Human  
Two-Month  
Combination

### PHASE III

Randomized  
Controlled Trial  
Efficacy

### CONFIRMATORY PROOF OF COMBINATION EFFICACY

**Big Gap**

## CRITICAL PATH DRUG DEVELOPMENT DECISIONS

*Which Models best inform critical decisions?*

Compound Selection / Regimen Evaluation

Early Indication of Efficacy of Individual Drugs and Limited Data on Combinations

Dose Selection / Regimen Evaluation

Reliability of Predictions  
Uncertain

Gold Standard for Confirmation of Efficacy (Durable Cure)

# CPTR Pre-Clinical and Clinical Sciences Workgroup

## Mission & Goals

### Mission

Develop and/or validate tools and innovative approaches to address pre-clinical issues including *in vitro* and *in vivo* efficacy, PKPD analyses using appropriate biomarkers, drug safety, metabolism, DDI, etc. These tools may be submitted to regulatory authorities for regulatory review and/or qualification as appropriate.

### Early goal related to pre-clinical *in vitro* and *in vivo* models

Evaluate the evidence base and develop criteria for evaluating the utility of various preclinical models to inform and test new drug regimens.

### Early Evidence

Landscape analysis\* identified HFS-TB as having an appropriate data inventory to assess predictive accuracy of a preclinical model for clinical outcomes.

# Early success

*EMA qualification opinion on the HFS-TB*

June 26, 2014

- HFS-TB qualified for use in drug development programs ***as additional and complementary tool***
- HFS-TB can be used in regulatory submissions, esp. for informed design and interpretation of clinical studies
- HFS-TB is recommended to be useful as follows:
  - To provide preliminary proof of concept for developing a specific drug or combination to treat tuberculosis
  - To select the pharmacodynamic target (e.g.  $T_{>MIC}$ , AUC/MIC)
  - To provide data to support PK/PD analyses leading to initial dose selection for non-clinical and clinical studies
  - To assist in confirming dose regimens for later clinical trials taking into account human PK data and exposure-response relationships

# Evaluation of *in vivo* models

*“Correlations between drug concentration and pathogen survival that are based on in vitro models cannot be expected to reiterate all aspects of in vivo antimycobacterial treatment.”*

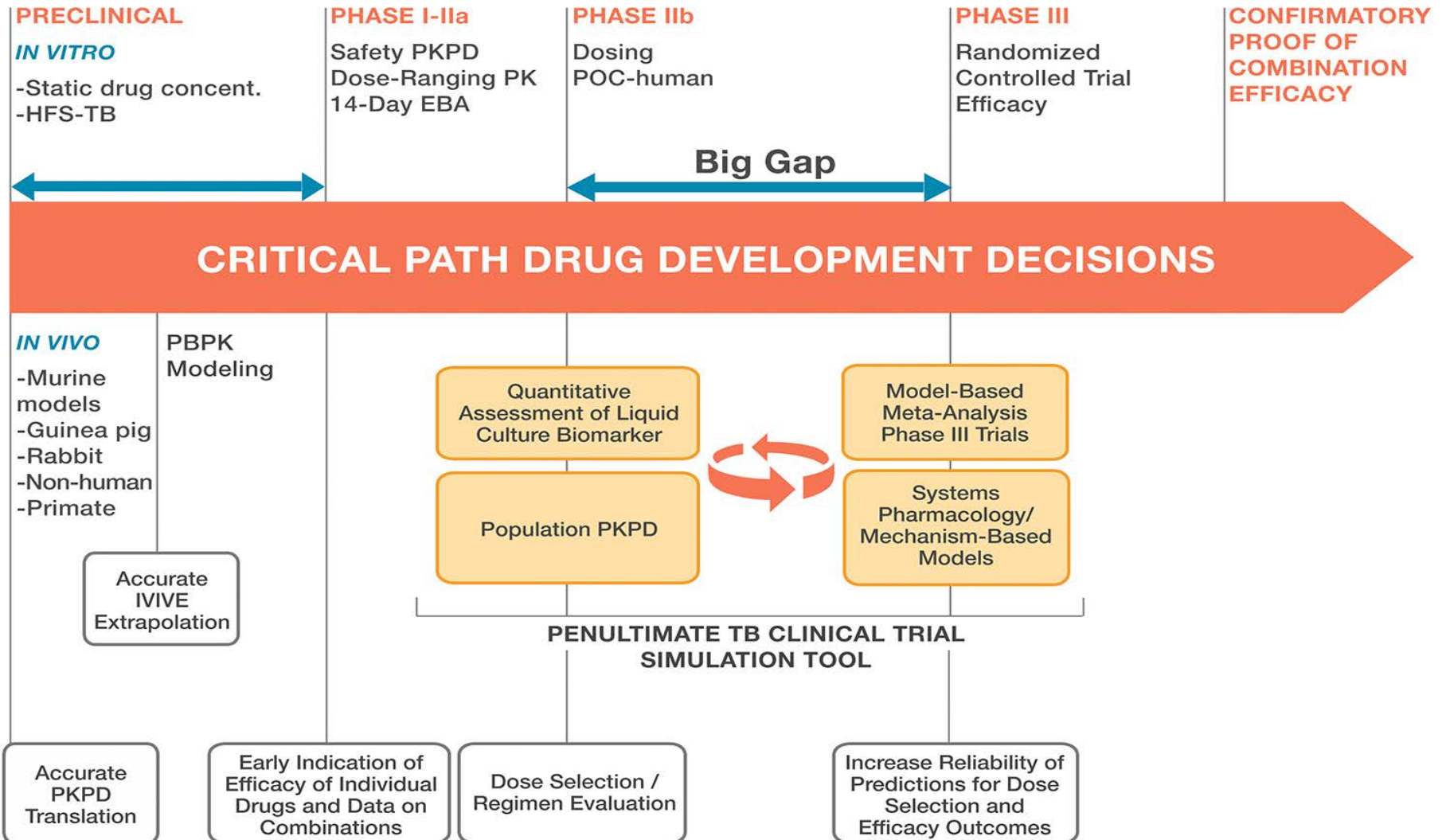
Chilukuri et al, CID 2015; 61(S1):S32

## Advantages of *in vivo* models

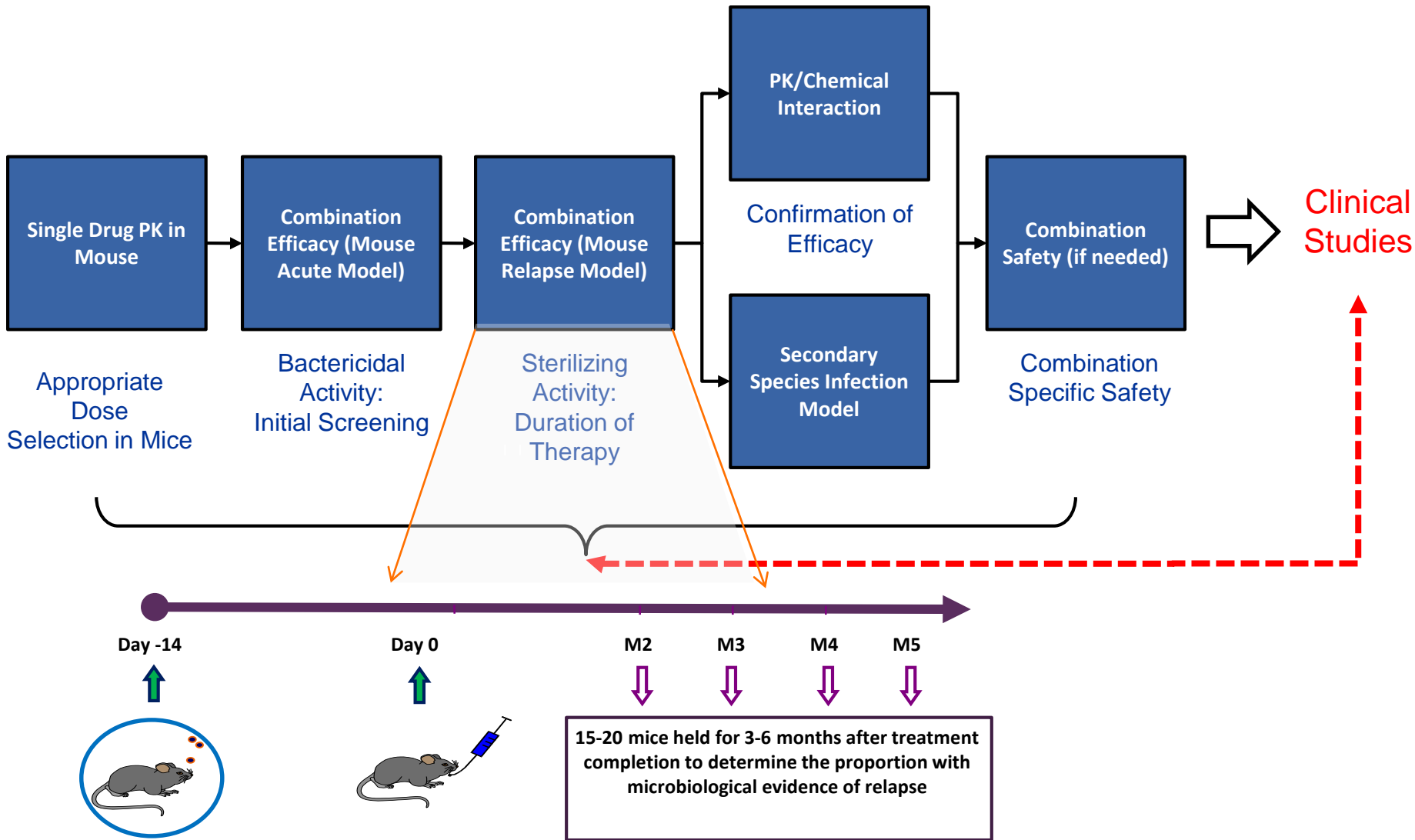
- Better reflect the phenotypic heterogeneity in bacterial populations as determined by host-pathogen interactions, including development of tissue pathology
- Present complexities of drug distribution to, and action at, various sites of infection

# Improving TB regimen development

## *Decreasing risk of attrition*



# Mouse model of sterilizing activity



# Evaluating the sterilizing mouse model

## Rationale

- Past and present role in TB regimen development
  - track record in forecasting treatment-shortening potential of RIF, PZA
  - relapse endpoint considered closest correlate of current phase 3 endpoint
- Amount of available data on regimens evaluated in clinical trials
- Does not preclude evaluation of other models
  - eg, C3HeB/FeJ mouse, marmoset



# Evaluating the sterilizing mouse model

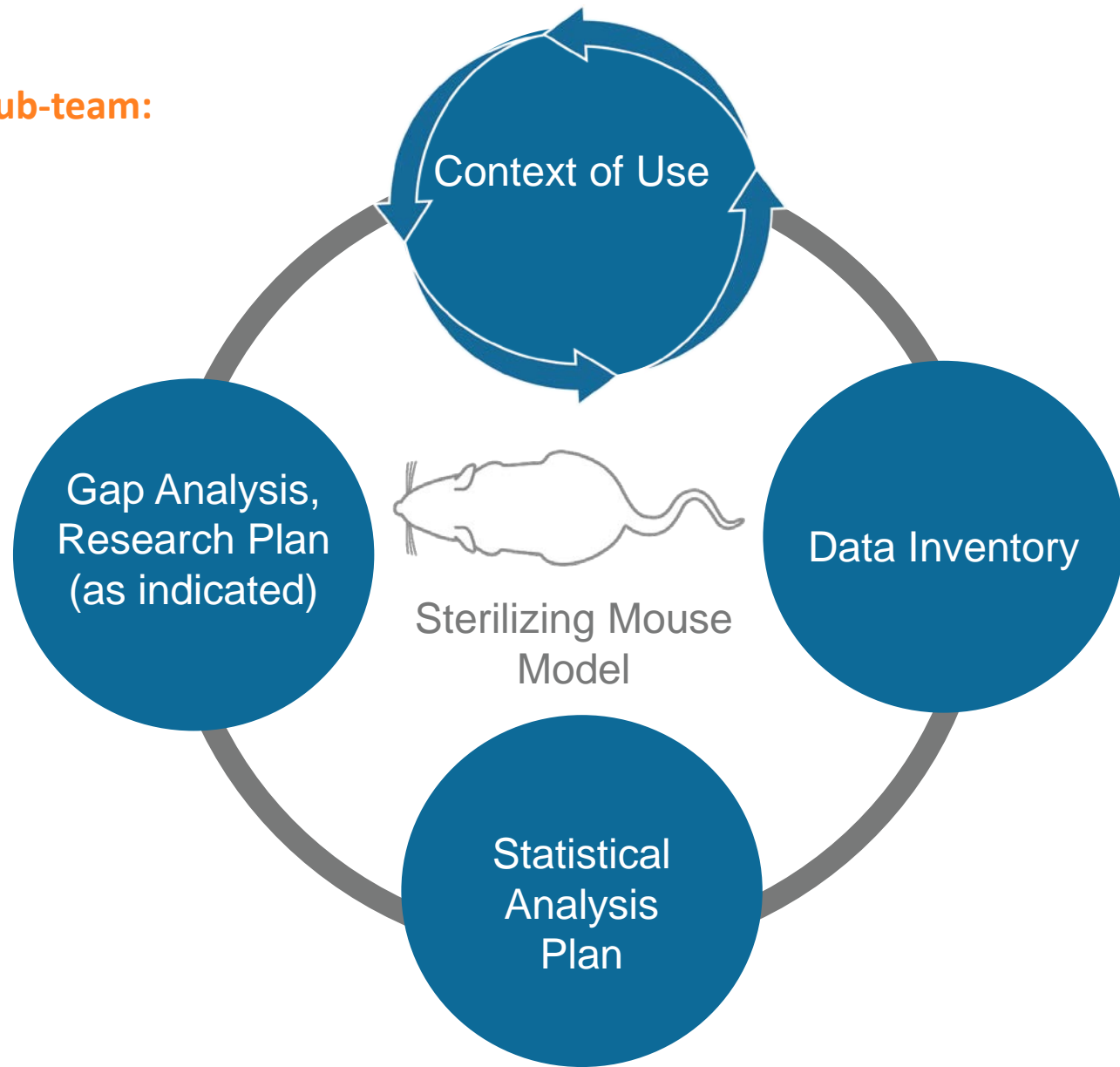
## General Aim

- Quantify the predictive accuracy of mouse models using relapse as an endpoint for the purpose of rank ordering regimens and estimating the effective treatment duration

# Workplan for evidence-based evaluation of sterilizing mouse model

## CPTR PCS-WG Mouse Model Sub-team:

Dr. Nicole Ammerman  
Dr. Al Berg  
Dr. Dakshina Chilukuri  
Dr. Geraint Davies  
Dr. Geo Derimanov  
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Dr. Mike Lyons  
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Dr. Eric Nuermberger  
Dr. Klaus Romero  
Dr. Rada Savic  
Dr. Christine Sizemore  
Dr. Peter Warner  
Lindsay Lehmann



# COU Scenario 1: Treatment Duration and Rank-Ordering of Regimens

## General Description

Experiments testing drug combinations in mice provide an additional and complementary tool to existing methodology to inform regimen selection, to maximize sterilizing effects.

Data produced will support submissions to regulatory agencies throughout the drug development process, to optimize design of clinical trials.

## Stage of Drug Development for Use

Non-clinical PK/PD testing

## Intended Application

The data from experiments in mice infected with *M. tuberculosis*, using relapse as the main endpoint, will be used to calculate treatment effect sizes, to then rank-order regimens and estimate clinical treatment duration.

# Data inventory

- Focus first on mouse strains other than C3HeB/FeJ (“Kramnik”)
- Inventory identified a variety of relapse-based pre-clinical studies with corresponding clinical trial outcomes data

Test regimen intervention	Regimen comparison	# of expts
Combining INH+STR	HS <u>vs.</u> H or S monotherapy	1
Shortening duration of INH+STR	6HS <u>vs.</u> 18HS	1
Adding RIF to INH+STR or INH+EMB+PZA	HR (or HRS or HREZ) <u>vs.</u> HS (or HEZ)	4
Adding STR to INH+RIF	HRS <u>vs.</u> HR	1
Adding PZA to INH+RIF ( $\pm$ STR/EMB)	HRZ (or HRSZ or HREZ) <u>vs.</u> HR (or HRS or HRE)	4
Shortening duration of PZA	2HREZ/4RH <u>vs.</u> 6HREZ	1
Increasing dose of RIF	High-dose R plus HEZ <u>vs.</u> HREZ	2
Extending dosing interval of 1 <sup>st</sup> -line Rx	HREZ (2/7) <u>vs.</u> HREZ (daily)	1
Replacing EMB with MXF	HRMZ <u>vs.</u> HRZ(E)	3
Replacing INH with MXF	MRZ(E) <u>vs.</u> HRZ(E)	10
Replacing RIF with RPT	HPZ(E) <u>vs.</u> HRZ(E)	7
Replacing RIF+EMB with RPT+MXF	HPMZ <u>vs.</u> HRZ	3
Replacing RIF with RPT and extending dosing interval (in continuation phase)	HP(1/7) cont phase <u>vs.</u> HR(2/7)	2
Comparing INH+RIF+PZA+EMB with PMD+MXF+PZA	PaMZ <u>vs.</u> HRZ(E)	8

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# Proposed statistical analysis plan

**An initial analysis approach using logistic regression based on relapse data is proposed. The following metrics will be obtained from the proposed analyses:**

- Covariate-adjusted probability of relapse for each regimen and duration
  - Can be used for rank order comparison
  - Absolute proportions can be compared across regimens and evaluated for correlations with human data when treated for the same duration
- Covariate-adjusted treatment duration required to achieve a specified relapse-free proportion (50% or 90%)
  - Can be used to rank order regimens on treatment shortening potential
  - Absolute differences between regimens can be compared to obtain the magnitude of treatment shortening relative to a control arm

**Subsequent analyses may examine bactericidal activity (CFU vs. time) and exposure-response**

# Lessons learned

- Overall, the amount of relevant preclinical and clinical data available are limited
  - few regimens are confirmed clinically to have different efficacious treatment durations
  - mouse expts not designed for precise delineation of treatment-shortening effects
  - little dose-ranging mouse data or human exposure-response analysis to examine the impact of variability in human PK, Mtb MIC
  - limited ability to explore potentially important covariates (eg, Mtb strain, route of infection, infectious dose, incubation period)
- As a categorical variable assessed monthly in individual mice, relapse is not as amenable to quantitative analysis as CFU cts
- Adopting a common data management system and/or common data elements could make such analyses simpler, more robust



# Summary points

- An initial step to address the “translational gap” is to learn what data from what models analyzed in what way best inform key trial design decisions.
- Evidence-based validation of pre-clinical models is important:
  - to confidently place preclinical models on the critical development path,
  - to increase the efficiency of regulatory interactions,
  - to set a precedent for objective, data-driven processes to apply to other models (e.g., C3HeB/FeJ mouse, marmoset), and
  - to identify gaps in knowledge & in existing tools to drive future research.
- Evaluation of sterilizing mouse models is the appropriate first step for *in vivo* models, with other models to follow

# Acknowledgements

## **CPTR PCS-WG Mouse Model Sub-team:**

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Dr. Peter Warner (Bill & Melinda Gates Foundation)

Lindsay Lehmann (Critical Path Institute)

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